RESEARCH

Open Access



Ultrasound screening protocol for osteochondrosis at selected predilection sites in thoroughbred yearlings

Seamus Hoey^{1*}, Jeremiah O'Sullivan², Jennifer Byrne¹, Sinead Devine¹, William Toomey², Hester McAllister¹ and Cliona Skelly¹

Abstract

Background: Osteochondrosis is a common condition of young horses where there is a failure of endochondral ossification, usually at predisposed sites. The estimated prevalence of osteochondrosis is 33–44%, with radiographic screening of yearlings being used to identify lesions. Radiography has two major limitations: poor sensitivity in detecting cartilaginous lesions and secondly, the exposure of the horse and personnel to ionising radiation. Ultrasonography allows imaging of the articular cartilage and subchondral bone margins and has been shown to be more sensitive in identifying osteochondrosis lesions. However, the ultrasonographic technique for examining joints is operator dependant, resulting in highly variable examinations, thus affecting its reliability and reproducibility as a screening test.

Results: A prospective observational clinical population study was undertaken involving twenty-two clinically normal weanling thoroughbred horses on-farm, describing a detailed protocol of the ultrasonographic examination technique for on-farm screening of common sites of osteochondral disease in the young horse, namely the carpal, metacarpophalangeal, stifle, tarsal and metatarsophalangeal joints.

Conclusion: Two veterinary practitioners used the technique to illustrate the repeatability of the protocol. The stepby-step protocol provides a valuable, reliable, repeatable technique for veterinary professionals performing screening ultrasound in the field.

Keywords: Equine, Joint, Osteochondrosis, Screening, Ultrasound

Background

Osteochondrosis is the most important developmental orthopaedic disease in horses [1] characterised by failure of endochondral ossification in the epiphyseal or metaphyseal growth plates, resulting in an area of thickened cartilage [2]. These areas may ossify and return to an almost normal condition. Alternatively, cartilage necrosis may develop resulting in a fissure

¹ Equine Clinical Studies, Diagnostic Imaging and Anaesthesia, School of Veterinary Medicine, University College Dublin, Dublin, Ireland Full list of author information is available at the end of the article

in the cartilage or subchondral bone, with subsequent cartilage flap formation, development of subchondral osseous cyst like lesions or collapse of the articular cartilage [3, 4]. With a shear stress, a cartilage flap fragment can develop. This fragment can detach leading to a free intraarticular body or "joint mouse", corresponding to osteochondrosis dissecans [5]. Osteochondrosis has a prevalence of 33–44% in horses, most frequently seen in Thoroughbreds and Warmblood horses [6–8]. Lesions may develop within the first seven months of age, but can be clinically silent until the horse is brought into work [9, 10]. In the weanling and yearling horses the most commonly



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visithttp://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence: seamus.hoey@ucd.ie

affected joints are the tarsocrural, femoropatellar, metacarpophalangeal and metatarsophalangeal joints, followed by the carpal, elbow, shoulder and cervical articular facet joints [11, 12].

A standardised set of screening "yearling radiographs" are acquired to highlight any abnormality which may affect the future racing and breeding potential [13]. Osteochondrosis lesions can negatively impact the price at sale [14, 15]. As part of the routine sales process each year, countless horses are exposed to ionising x-ray radiation, and the teams of personnel acquiring the images are also exposed to scatter radiation [16].

Equine joint ultrasound examinations are usually focused in nature where lameness is localised to a joint or set of joints. The common use of ultrasound in general practice facilitates the majority of equine practitioners have access to ultrasound equipment, with good quality, reasonably priced, mobile ultrasound machines [17].

Ultrasonographically, articular cartilage is a smooth hypoechoic to anechoic band, superficial to a smooth curvilinear hyperechoic surface eliciting distal acoustic shadowing, consistent with the cartilage-subchondral bone interface [18]. Several authors describe the evaluation of the periarticular and articular anatomy of the joints, as well as specific examples of disease [2, 18–32]. Osteochondrosis can manifest as an interruption in the subchondral osseous margin, depressions or semi-circular indentations in the cartilaginous or subchondral interface [17, 22]. However, ultrasound technique is very operator dependant and interpretation of the images acquired may be difficult when assessed by a reviewer other than the ultrasonographer.

The aim of this study is to develop a step- by step ultrasound protocol for on-farm screening of horses for the most common sites of osteochondrosis that can be performed by veterinary practitioners and produce standardised images.

Results

Twenty Two weanling Thoroughbred horses with a mean age of 314 days (range of 281–367 days) were included in the study, 11 male and 11 female. Ultrasonographic studies were performed in less than one hour (15 to 58 min). With practice, the time frame for completing the ultrasound examination was between 15 and 35 min.

Administration of ample amounts of warmed alcohol facilitated good image quality despite the unclipped hair. The articular cartilage and subchondral bone margin were readily visualised. Frequently a thin hyperechoic line was identified superficial to the hypoechoic cartilage, representing the cartilage-synovial fluid interface.

Carpal joints

The extensor carpi radialis was easily identified at the mid-cranial aspect of the distal radial physis (Fig. 1). The hyperechoic margins of the carpal bones were smooth and well defined. The intercarpal joint spaces were easily visualised with well-defined intercarpal ligaments (Fig. 2). The antebrachiocarpal joint (ABCJ), middle carpal joint (MCJ) and carpometacarpal (CMCJ) margins are sharp, with narrow hypoechoic interfaces. The adjacent radial,







carpal, and metacarpal bones were well defined (Fig. 3). Mild smooth hyperechoic projections from the articular margins of the carpal bones were identified, with distal acoustic shadowing and normal adjacent articular margins of the opposing bone (Fig. 4). Minimal irregular undulation of the hyperechoic osseous margins was seen at the dorsoproximal extent of the carpal bones on transverse images, with the third carpal bone (C3) most affected (Fig. 5).

Metacarpophalangeal/metatarsophalangeal joints

The metacarpophalangeal (MCPJ) and metatarsophalangeal joints (MTPJ) were the most technically challenging joints to evaluate, in both standing and partial flexion. Standing, the dorsal margin of the sagittal ridge of the distal third metacarpal/metatarsal bone (MC3/MT3) showed well-defined cartilage and subchondral bone margins (Fig. 6). The dorsodistal margin of the sagittal ridge frequently showed irregular subchondral bone





margination, with hypoechoic linear to irregular echogenicities (compared to the subchondral bone margin) deep to the normal articular cartilage margins (Fig. 7). In partial flexion the sagittal ridge was more ill-defined when imaged through the intersesamoidean ligament (Figs. 8 and 9). Mild undulation and irregularity of the osseous margins of the sesamoid bones was frequently identified. Mild indentations of the hyperechoic interface were seen with and without focal areas of hypoechogenicity of the adjacent attachment of the branches of the tendon of the third interosseous medius muscle (previously referred to as the suspensory ligament) (Fig. 10). The palmaroproximal aspects of the palmar eminences of the proximal phalanx (P_1) were imaged by minimal fanning of the transducer in a palmaroproximal-dorsodistal direction (Fig. 11).

Stifle joint

The femoral trochlear ridges (TRs) were well defined and smoothly marginated showing hyperechoic







subchondral bone and hypoechoic superficial cartilage (Figs. 12 and 13). There was mild blunting of the craniodistal subchondral bone margin of the medial femoral TR, with equivocal heterogeneity of the superficial cartilage layer (Fig. 14). The femoral and tibial condyles are well defined, surrounding the menisci (Fig. 15).

Tarsal joints

The TRs of the talus showed well-defined smoothly curved subchondral bone margins (Fig. 16). Mild indentation of the subchondral bone and thinning of the superficial cartilage was identified intermittently at the cranioproximal aspect of the medial femoral TR, without any evidence of joint effusion or irregular margins,





likely a normal variant (Fig. 17). The central and distal rows of tarsal bones showed well-defined articular margins (Fig. 18). The well-defined dorsal protuberance of the third tarsal bone showed a smooth extension of the hyperechoic osseous margin (Fig. 19). The distal intermediate ridge of the tibia (DIRT) was clearly visible (Fig. 20), with mild proximomedial to distolateral rotation necessary due to the oblique orientation of the TRs of the talus (Fig. 21). The longitudinal view of the medial malleolus required fine proximomedial to distolateral rotation and rocking of the transducer. There was some undulation of the medial margin of the medial malleolus in several horses (Fig. 22).

Screening protocol pilot

One veterinary practitioner; sonographer 2, had 20 years of clinical experience and felt moderately experienced in ultrasound and mildly experienced in equine







joint ultrasound. Sonographer 2 reported that the technique was relatively easy to perform and spent 40 min and 22 s on the first examination without stifle flexion, and 38 min and 58 s on the second examination with stifle flexion.

The second practitioner; sonographer 3, had 2.5 years of experience and reported being relatively inexperienced in ultrasound. Sonographer 3 reported that the technique was somewhat difficult to perform and spent 45 min and 22 s on the first examination without stifle flexion, and 39 min and 49 s on the second examination with stifle flexion.

On specific joints, both reported that images of the carpus, metacarpophalangeal (in flexion and extension), metatarsophalangeal (in flexion and extension) and stifle (in flexion and extension) joints were relatively easy to very easy to acquire. Both observers reported that the tarsus was relatively difficult, with both reporting that the speed and difficulty of image acquisition improved with practice.





Discussion

A systematic protocol of the ultrasonographic assessment of a selection of the most common sites of osteochondrosis in yearling horses is described in detail, facilitating the use of the protocol by veterinary practitioners with variable experience.

Radiography is currently the recognised gold standard for equine orthopaedic screening, particularly in the case of pre purchase examinations. A major limitation of radiography is the inability to distinguish cartilage from the other soft tissue opacities of the joint. In radiography, the deformation of the subchondral bone margins, change in opacity of the subchondral bone and soft tissue swelling are signs suggestive of osteochondral disease. Subtle cases involving only the articular cartilage may be missed.

Ultrasound can directly image cartilage and the subchondral bone margin, and without exposure to





ionising radiation. Ultrasonographic detection of osteochondrosis of the canine shoulder and the equine tarsocrural joints have been described [33], and show a higher sensitivity in the detection of osteochondrosis, compared to radiography [23, 34]. In humans, ultrasound screening for osteochondrosis lesions in young athletes describe positive predictive values of 66.7 to 100%, when compared to MRI. Indeed, the measurement of defects identified ultrasonographically were similar to those of MRI [35, 36]. All ultrasound examinations of the weanling thoroughbred horses performed in early spring. The haircoat at this time was not very long or dense, and the fine hair and skin of the thoroughbred weanling permitted adequate ultrasound beam penetration and quality image acquisition. In some cases, the area of interest was brushed to remove dirt or debris from the hair and to allow direct alcohol application to the skin and hair. In some cases, repeat application of alcohol was necessary for optimal imaging, which was well tolerated by the horse. It was not





possible to clip the joints of these horses for the examinations, and in clinical practice it would not be feasible to clip each horse for ultrasound on aesthetic grounds. Ultimately all joints in all horses were imaged fully with good image quality yielding comparable, repeatable images.

Despite the ubiquitous presence of ultrasound in general equine practice, there is little literature describing a standardised technique in joint imaging. Many texts describe the normal ultrasonographic anatomy of joints, with an emphasis on the soft tissue structures such as periarticular tendons, ligaments, muscles, tendon sheaths and bursae as well as the joint capsule and synovium. The specific ultrasound technique is rarely or incompletely described [2, 20, 25–31, 37, 38]. The inconsistency in ultrasound technique results in images that are often not comparable with follow up images, or that the images can't be reliably assessed by anyone other than the sonographer. In radiography there is a defined collection of radiographic views acquired for optimal assessment of the yearling horse at the time of sale. These views are commonly performed in specialty and first opinion practice and are thus repeatable and comparable tests.





In ultrasound examinations, the skill of the ultrasonographer in the handling and manipulation of the ultrasound transducer can vary greatly. Images recorded in an ultrasound examination can be variable in the anatomy imaged, the imaging plane/transducer orientation and the ultrasound imaging settings (such as focal zone placement, frequency, transducer used, gain settings). This limits the repeatability and comparability of the images between horses, and between ultrasonographers. By describing a detailed protocol indicating optimal transducer orientation and movements, the imaging planes can be standardised facilitating repeatable and comparable image recording. In this way, recorded images can be subsequently reviewed and compared between horses and between time points.

The initial decision to remove the flexed stifle for the additional observers was the perception that imaging the flexed stifle in a yearling would be difficult and time consuming. The observers did not perceive the flexed stifle as a more difficult component of the examination, and the visualisation of the more caudodistal aspects of the femoral condyles would be important in the search



Marker is to proximal



for osteochondral disease. Both observers noted that imaging the tarsus was more challenging reflecting the more complex anatomy. However, confidence improved with practice, and both observers acquired diagnostic images. Both practioners highlighted the importance of having reference images to emulate ensuring the images acquired were consistent and repeatable.

A limitation of ultrasound is that the ultrasound waves are unable to penetrate bone, and as a result restrict ultrasonographic assessment of deeper structures. In this study movement and flexion of the stifle and metacarpophalangeal joints were used to better interrogate deeper articular margins. Despite this limitation, most common sites of osteochondrosis of the fore and hindlimb joints can be thoroughly accessed over the course of the examination. Diagnostic images could be acquired through the use of warmed alcohol as a couplant placed directly on haired skin. By removing the need to clip these weanlings, the screening process is more attractive to owners and trainers.



The images acquired of the clinically normal weanling horses showed multiple repeatable echogenic changes within the trochlear ridges of the femur and talus, and within the sagittal ridge of the metacarpal and metatarsal bones. These changes were not associated with joint effusion or swelling. Futher study would be required to compare the ultrasonographic findings to the radiographic findings at the time of ultrasound, and to the radiographic findings at the time of yearling radiographic imaging.

The technique does not include all the sites of disease in the juvenile horse but focuses on the most common sites of osteochondral disease. By moving through the imaging planes, cineloops can be recorded, documenting the most common sites of osteochondral disease. The imaging planes would include the osteochondral sites of osteochondrosis as well as the dorsal aspects of the joints, where osteochondral (OCD) fragments may be identified. The site of origin of osteochondrosis may be identified, where identification of free osteochondral fragments may not be captured in the imaging planes, a diagnosis of osteochondrosis would remain. These cineloop images can be reviewed by the ultrasonographer as well as another observer, where images are repeatable and comparable. Further studies are warranted, comparing the ultrasound protocol with contemporaneous radiographic examinations. The ultrasound examination protocol may be useful as an initial screening test, where any equivocal findings could be further examined using radiography. The screening protocol would not replace a diagnostic examination targeted at a specific site of lameness or pain.

Conclusion

An ultrasonographic screening protocol is described in a detailed systematic manner facilitating on-farm screening using mobile ultrasound equipment. The protocol has been shown to be feasible for practitioners of varied experience in ultrasound, thus can have a role as a potential screening method capable of reproducing standardised images, in a reasonable time frame with only minimal practice.

Materials and methods

Thoroughbred weanlings were included as part of the annual screening programme on three farms. A clinical examination of each weanling demonstrated normal gait at walk and normal cardiac auscultation. Light sedation using intravenous administration of 3.5 mg of Detomidine (Domosedan, Zoetis) 3.5 mg of Butorphanol (Torbugesic, Zoetis) and 3.75–5 mg of Acepromazine (Calmivet, Vetoquinol).

The ultrasonographic examinations were performed by an ECVDI/ACVR (EDI) board certified veterinary radiologist. The hair and skin of each joint was thoroughly soaked with warmed alcohol. A portable ultrasound machine (GE Logiq R7) and a 5–13 MHz Linear Transducer (GE 12L-RS) was used with focal zone(s), frequency, time gain compensation (TGC) and overall gain optimised by the ultrasonographer. The transducer was held with the marker located medially in a transverse orientation or proximally in the longitudinal plane. Representative cineloop images were recorded and saved for each weanling, facilitating subsequent review.

Protocol

The order examination was left forelimb, left hindlimb, right hindlimb, and right forelimb. This order allowed optimal sedation during the hindlimb examinations.

Carpus (Fig. 23)

Three transverse oriented imaging planes are obtained.

i. In transverse, identify the extensor carpi radialis tendon (ECRT) dorsal to the distal radial physis. Slide distally, imaging the ABCJ, intermediate carpal bone, MCJ, intermediate facet of C_3 , CMCJ, and proximal MC3.

ii. Return to the ECRT in transverse, at the distal physis of the radius. Slide medially until the tendon is on the lateral aspect of the image. Slide distally imaging the ABCJ, radial carpal bone, MCJ, radial facet of C_{3} , CMCJ and proximal MC2.

iii. Return to the ECRT in transverse orientation, at the distal physis of the radius. Slide lateral to position the tendon on the medial aspect of the image. Slide distally imaging the ulnar carpal bone, fourth carpal bone and proximal MC4.

Two longitudinal oriented imaging planes are obtained.

iv. Rotate 90°into longitudinal plane. Starting at the medial aspect of the ABCJ, slide laterally across the dorsal surface of the ABCJ.

v. From lateral, slide distally to the lateral aspect of the MCJ. Slide medially imaging the dorsal surfaces of the MCJ and CMCJ.

Metacarpophalangeal joint (Fig. 24)

Three transverse imaging planes are obtained dorsally.

i. Begin in transverse, at the dorso proximal aspect of the MC_3 sagittal ridge, at the chondro-osseous margin. Slide distally imaging the sagittal ridge, MCPJ and the P_1 .

ii. Return to midline of the distal MC_{3} , to the level of the chondo-osseous margin. Slide laterally to the lateral aspect of the chondro-osseous margin. From there, slide distally imaging the entire lateral condyle.

iii. Return to the distal MC_3 , to the level of the chondo-osseous margin. Slide medially to the medial aspect of the chondro-osseous margin. From there, slide distally to image the medial condyle.

One longitudinal imaging plane is obtained dorsally.

iv. Return to the distal MC_3 chondro-osseous margin beginning at the medial aspect, and rotate 90°into longitudinal orientation. From here slide laterally to image the medial and lateral aspects of the MC_3 condyle, the dorsoproximal margin of the P_1 and the dorsal margin of the MCPJ.

One longitudinal and one transverse plane are obtained of the branches of the tendon of the third interosseous medius muscle (suspensory ligament) to the palmar eminences of P_1 .

v. In longitudinal, slide distally imaging the entire lateral branch of the tendon of the third interosseous medius muscle (suspensory ligament) to its insertion on the lateral sesamoid bone. Continue sliding distally along the lateral sesamoid to the lateral palmar eminence of P_1 .

vi. Return to the distal aspect of the lateral branch of the tendon of the third interosseous medius muscle







in longitudinal plane extending in a lateromedial direction. The paths i-ix are described in the text

(suspensory ligament) and rotate 90° into transverse plane. Slide distally imaging the lateral sesamoid and the lateral palmar eminence of P₁.

Repeat the process on the medial aspect of the limb.

Two transverse planes and one sagittal plane are obtained with partial limb flexion.

Lift the foot, flexing the MCPJ.

vii. Begin in transverse at the palmar midline of the fetlock, imaging the palmaroabaxial margins of the medial and lateral sesamoids and the Intersesamoidean ligament. Using the intersesamoidean ligament as a window, evaluate the accessible portion of the palmar sagittal ridge of MC3. Sliding distally, image the palmaroproximal aspect of P_1 . viii. Remaining in flexion, return the transducer, in transverse orientation, to the dorsal mid-line surface of the fetlock imaging the dorsoproximal aspect of the sagittal ridge of MC_3 . Slide from proximal to distal to assess the sagittal ridge and dorsoproximal P_1 .

ix. Return to the dorsomedial aspect of the distal MC_3 and rotate 90° into longitudinal orientation. Beginning medially, slide dorsally and laterally to image the sagittal ridge of the MC_3 , the dorsoproximal margin of the P_1 and the dorsal margin of the MCPJ.

Stifle joints (Fig. 25)

Two transverse planes are imaged of the femoral TRs and condyles.

In transverse identify the cranial ridge of the patella at its craniolateral aspect.

i. Slide laterally placing the cranial ridge of the patella to the medial aspect of the image. Slide distally to image the lateral femoral TR.

ii. Return to the patella and slide medially, placing the cranial ridge of the patella to the lateral aspect of the image. Slide distally to image the medial femoral TR.

Two longitudinal planes are imaged.

direction. The paths i-vi are described in the text

iii. Rotate into a longitudinal plane. Place the transducer at the medial aspect of the femorotibial joint to identify the medial meniscus. Slide slightly proximal to image the medial femoral condylar margin and the recess of the femorotibial joint. From this point, slide cranially, imaging the medial femoral condyle and TR. If the entire femoral trochlear ridges are not included in the imaging plane, a subsequent examination may be repeated placing the transducer more proximally.

iv. Continue to slide laterally to the extensor fossa at the abaxial aspect of the lateral femoral condylar surface. Slide craniodistally approximately 2 cm and rotate approximately 45° in a cranioproximal-caudodistal orientation, to better identify the lateral meniscus, and adjacent femoral and proximal tibial condyles. Slide cranially



of the stifle joint. Images are acquired in transverse plane extending in a proximal to distal direction, and in longitudinal plane extending in a lateromedial

iv

iii



Fig. 26 Lateromedial and plantarodorsal views of the tarsus, showing the paths of the ultrasound transducer in the examination of the tarsal joints. *Images are acquired in transverse plane extending in a proximal to distal direction, and in longitudinal plane extending in a lateromedial direction. The paths i-v are described in the text*

and medially imaging the lateral femoral condyle and lateral TR and proximal tibial margins.

One transverse plane is imaged in flexion.

The hindlimb is lifted to flex the stifle. A second person, or a hoof stand may be necessary.

v. In transverse orientation, begin at the distomedial aspect of the femur, at the proximal margin of the medial femoral TR. Slide distally to image the medial femoral TR.

vi. Repeat for the lateral femoral TR. At the distal-most aspect the beam is angled caudoproximally to include as much of the condylar surfaces as possible.

Tarsus (Fig. 26)

Three transverse imaging planes are obtained.

i. In transverse, at the craniodistal midline aspect of the tibia, identify the fibularis tertius tendon and slide distally to image the DIRT. The transducer may require proximomedial to distolateral rotation between the TRs of the talus to fully identify the DIRT.

ii. Remaining in transverse, slide laterally to the centre of the lateral TR. Slide distally to assess the lateral talus TR.

iii. Return to the DIRT in transverse plane, sliding medially to centre on the medial talus TR. Slide distally to assess the medial talus TR.

Two longitudinal imaging planes are obtained.

iv. Rotate 90° into longitudinal, slide to the medial aspect of the tarsocrural joint, to the medial malleolus, at the articulation with the medial trochlear ridge of the talus. A slight proximomedial to distolateral transducer

rotation may improve surface contact. From here slide laterally imaging the medial TR, DIRT and lateral TR. Mild proximal rocking of the transducer helps to better assess the margins of the DIRT.

v. Slide plantarodistally to the plantar tubercle of the fourth tarsal bone. From this point, slide medially imaging the proximal intertarsal, distal intertarsal and tarsometatarsal joints.

Metatarsophalangeal joint

As the metacarpophalangeal joint.

Screening protocol pilot

Two equine veterinary practitioners were recruited. The practitioners performed the examination protocol twice; immediately after viewing a complete protocol demonstration and again six weeks later. Examination duration and images were recorded.

The initial examination did not include the flexed stifle, subsequently included in the second examination. Each observer completed a questionnaire (see supplemental information) relating to their previous ultrasound experience and the difficulty of the examination.

Abbreviations

ECRT: Extensor carpi radialis tendon; ABCJ: Antebrachiocarpal joint; MCJ: Middle carpal joint; C3: Third carpal bone; CMCJ: Carpometacarpal joint; MCPJ: Metacarpophalangeal joint; P1: Proximal phalanx; MC3: Third metacarpal bone; MC2: Second metacarpal bone; MC4: Fourth metacarpal bone; TR: Trochlear ridge; DIRT: Distal intermediate ridge of the tibia.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13620-022-00216-7.

Additional file 1.

Authors' contributions

SH, JOS, HMcA, CS-conceptualisation of the study and supervision of the project. SH, JOS, HMcA, CS- design of the methodology. SH-investigation of the literature. SH, JOS, WT, JB, SD-project fieldwork. SH, JOS, HMcA, CS-writing of the manuscript. SH, JOS, WT, JB, SD, HMcA, CS-revision of the manuscript. All authors read and approved the final manuscript.

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Declarations

Ethics approval and consent to participate

The study was performed with the ethical approval of the Institutional Animal Research Ethics Committee, University College Dublin (AREC-E-20-17-Hoey).

Competing interests

The authors declare that they have no competing interests.

Author details

¹Equine Clinical Studies, Diagnostic Imaging and Anaesthesia, School of Veterinary Medicine, University College Dublin, Dublin, Ireland. ²Clinic na gCapall, Farranacurragh, Oldleighin, Co. Carlow, Ireland.

Received: 21 January 2022 Accepted: 14 April 2022 Published online: 27 April 2022

References

- Mcllwraith CW. Osteocondrosis. In: Baxter GM, editor. Adams and Stashak's lameness in horses. 6th ed. Chichester: Wiley-Blackwell; 2011. p. 1155–65.
- Bourzac C, Alexander K, Rossier Y, Laverty S. Comparison of radiography and ultrasonography for the diagnosis of osteochondritis dissecans in the equine femoropatellar joint. Equine Vet J. 2009;41(7):685–92.
- van Weeren PR. 5 osteochondritis Dissecans. In: McIlwraith CW, Frisbie DD, Kawcak CE, van Weeren PR, editors. Joint disease in the horse. 2nd ed. Edinburgh: W.B. Saunders; 2016. p. 57–84.
- Firth EC, Weeren PR, Pfeiffer DU, Delahunt J, Barneveld A. Effect of age, exercise and growth rate on bone mineral density (BMD) in third carpal bone and distal radius of Dutch warmblood foals with osteochondrosis. Equine Vet J. 1999;31(S31):74–8.
- Laverty S, Girard C. Pathogenesis of epiphyseal osteochondrosis. Vet J. 2013;197(1):3–12.
- Mendoza L, Lejeune JP, Caudron I, Detilleux J, Sandersen C, Deliege B, et al. Impact of feeding and housing on the development of osteochondrosis in foals-a longitudinal study. Prev Vet Med. 2016;127:10–4.
- Vos NJ. Incidence of osteochondrosis (dissecans) in Dutch warmblood horses presented for pre-purchase examination. Ir Vet J. 2008;61(1):33.
- Vander Heyden L, Lejeune JP, Caudron I, Detilleux J, Sandersen C, Chavatte P, et al. Association of breeding conditions with prevalence of osteochondrosis in foals. Vet Rec. 2013;172(3):68.
- Sandgren B, Dalin G, Carlsten J. Osteochondrosis in the tarsocrural joint and osteochondral fragments in the fetlock joints in Standardbred trotters. Epidemiol Equine Vet J. 1993;25(S16):31–7.
- 10. Walmsley JP. Diagnosis and treatment of ligamentous and meniscal injuries in the equine stifle. Vet Clin N Am Equine Pract. 2005;21(3):651–72.
- 11. Ortved KF. Surgical Management of Osteochondrosis in foals. Vet Clin North Am Equine Pract. 2017;33(2):379–96.

- 12. van Weeren PR, Barneveld A. The effect of exercise on the distribution and manifestation of osteochondrotic lesions in the warmblood foal. Equine Vet J Suppl. 1999;31(31):16–25.
- Martin BB, Kimmel JC, Cheney MW, Ellis DR, Ross MW. The sales yearling. In: Ross MW, Dyson SJ, editors. Diagnosis and Management of Lameness in the horse. Saint Louis: W.B. Saunders; 2003. p. 836–41.
- Mcllwraith CW. Inferences from referred clinical cases of osteochondritis dissecans. Equine Vet J. 1993;25(S16):27–30.
- Verwilghen DR, Janssens S, Busoni V, Pille F, Johnston C, Serteyn D. Do developmental orthopaedic disorders influence future jumping performances in warmblood stallions? Equine Vet J. 2013;45(5):578–81.
- Tyson R, Smiley DC, Pleasant RS, Daniel GB. Estimated operator exposure for hand holding portable X-ray units during imaging of the equine distal extremity. Vet Radiol Ultrasound. 2011;52(2):121–4.
- Martel G, Crowley D, Olive J, Halley J, Laverty S. Ultrasonographic screening for subclinical osteochondrosis of the femoral trochlea in foals (28-166 days old): a prospective farm study. Equine Vet J. 2018;50(3):312–20.
- Tomlinson JE, Redding WR, Sage A. Ultrasonographic evaluation of tarsocrural joint cartilage in normal adult horses. Vet Radiol Ultrasound. 2000;41(5):457–60.
- 19. Penninck DG, Nyland TG, O'Brien TR, Wheat JD, Berry CR. Ultrasonography of the equine stifle. Vet Radiol. 1990;31(6):293–8.
- 20. Adrian AM, Barrett MF, Werpy NM, Kawcak CE, Chapman PL, Goodrich LR. A comparison of arthroscopy to ultrasonography for identification of pathology of the equine stifle. Equine Vet J. 2017;49(3):314–21.
- 21. Cauvin ER, Munroe GA, Boyd JS, Paterson C. Ultrasonographic examination of the femorotibial articulation in horses: imaging of the cranial and caudal aspects. Equine Vet J. 1996;28(4):285–96.
- 22. Denoix J-M. Ultrasonographic examination of joints, a revolution in equine locomotor pathology. Bulletin de l'Académie vétérinaire de France. 2009;192(1):521–40.
- Relave F, Meulyzer M, Alexander K, Beauchamp G, Marcoux M. Comparison of radiography and ultrasonography to detect osteochondrosis lesions in the tarsocrural joint: a prospective study. Equine Vet J. 2009;41(1):34–40.
- 24. Rottensteiner U, Palm F, Kofler J. Ultrasonographic evaluation of the coxofemoral joint region in young foals. Vet J. 2012;191(2):193–8.
- Tnibar M, Kaser-Hotz B, Auer JA. Ultrasonography of the dorsal and lateral aspects of the equine carpus: technique and Normal appearance. Vet Radiol Ultrasound. 1993;34(6):413–25.
- Cauvin ERJ, Smith RKW. Ultrasonography of the fetlock. In: Kidd JA, Lu KG, Frazer ML, editors. Atlas of equine ultrasonography. Chichester: John Wiley & Sons; 2014. p. 45–72.
- Cauvin ERJ. Ultrasonography of the stifle. In: Kidd JA, Lu KG, Frazer ML, editors. Atlas of equine ultrasonography. Chichester: John Wiley & Sons; 2014. p. 161–81.
- Carstens A. Ultrasonography of the carpus. In: Kidd JA, Lu KG, Frazer ML, editors. Atlas of equine ultrasonography. Chichester: John Wiley & Sons; 2014. p. 107–23.
- Garrett KS. Ultrasonography of the hock. In: Kidd JA, Lu KG, Frazer ML, editors. Atlas of equine ultrasonography. Chichester: John Wiley & Sons; 2014. p. 149–59.
- Reef VB. Equine diagnostic ultrasound. Philadelphia: W B Saunders Company; 1998. xv + 560 pp.
- Reef VB, Whittier M, Allam LG. Joint ultrasonography. Clin Tech Equine Pract. 2004;3(3):256–67.
- Denoix JM, Audigie FA. Ultrasonographic Examinations of Joints in horses. San Diego: American Association of Equine Practitioners; 2001. p. 366–75.
- 33. Vandevelde B, Van Ryssen B, Saunders JH, Kramer M, Van Bree H. Comparison of the ultrasonographic appearance of osteochondrosislesions in the canine shoulder with radiography, arthrography, and arthroscopy. Vet Radiol Ultrasound. 2006;47(2):174–84.
- Wall CR, Cook CR, Cook JL. Diagnostic sensitivity of radiography, ultrasonography, and magnetic resonance imaging for detecting shoulder osteochondrosis/osteochondritis dissecans in dogs. Vet Radiol Ultrasound. 2015;56(1):3–11.
- Jungesblut OD, Berger-Groch J, Meenen NM, Stuecker R, Rupprecht M. Validity of ultrasound compared with magnetic resonance imaging in evaluation of osteochondritis Dissecans of the distal femur in children. Cartilage. 2019;12(2):194760351982843.

- Han K-J, Kim Y-K, Lim S-K, Park J-Y, Oh K-S. The effect of physical characteristics and field position on the shoulder and elbow injuries of 490 baseball players: confirmation of diagnosis by. Magn Reson Imaging. 2009;19(4):271–6.
- 37. Smith M, Smith R. Diagnostic ultrasound of the limb joints, muscle and bone in horses. Practice. 2008;30(3):152–9.
- M. H, J. S. A standardized protocol for the ultrasound examination of the equine stifle. Vet Radiol Ultrasound. 2004;45(6):594.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

